Role of the Predaceous Insects in Regulating the Population of the Main Piercing-Sucking Insect Pests Attacking Certain Vegetable Crops

T. E. Ata

Plant Protection Department, Faculty of Agriculture, Damietta University, Egypt.

*Corresponding author E-mail: drtarekata@du.edu.eg (T. Ata)

ABSTRACT:

The experiments were conducted to study the role of predaceous insects in regulating the population of vegetable crops insect pests at Kafr-Saad area, Damietta Governorate, Egypt during summer plantation of 2022. Nine predators belonging to four insect orders; i.e. Copleoptrea, Heteroptera, Diptera and Nuroptera were recorded on cucumber, cowpea, tomato and sweet potato. The coccinellid predators (*Coccinella undecimpunctata, C. septempunctata, Cryptolemus montiziri* and *Chilocorus nigritus*) were more abundant species on the tested host plants than the other predator species; *Orius* spp., *Macrolophus* sp., *Aphidoletes aphidimyza, Syrphus* sp. and *Chrysoperla carnea*. The coccinellid predators represented by 26.8, 29.7, 29.9 and 26.7% of the total number of predaceous insects in cucumber, cowpea, tomato and sweet potato crops, respectively. While, *Syrphus* sp. recorded the smaller number and represented by 5.4, 9.2, 11.2 and 10.6% of the total numbers of predatory insects on the previous crops. In respect to the preference of predators for the host plants, all predators greatly prefer cucumber (4504 predators / 25 leaves) followed by cowpea (757 predators / 25 leaves) and tomato (411 predators / 25 leaves) while, sweet potato plants showed a less preference for all predatory insects and represented by (292 predators / 25 leaves).

Keywords: Predatory insects, Vegetable crops, Piercing-Sucking Insects.

INTRODUCTION

Vegetables are very important for human nutrition; around 200 countries cultivate vegetables, which are a food source for people in many regions of the world, especially as providers of vitamins (C, A, B1, B6, B9, E), minerals, dietary fiber, and phytochemicals (Wargovich, 2000). Cucumber (Cucumis sativus L.) is one of the most important fresh vegetables consumed worldwide. In Egypt it is recently considered one of the most important vegetables grown in greenhouses and open field. The total cultivated area of open field cucumbers in 2018/2019 was 52.67 thousand acres and produced about 496.81 thousand tons of fresh fruits. The grain legume, cowpea (Vigna unguiculata L. Walp) is one of the most significant vegetables in Egypt. According to Belane and Dakora (2009), cowpea grain contains 23% protein and 57% carbohydrates, whereas the leaves contains 27-34% protein. Additionally, cowpea is a significant source of soil nitrogen, particularly in regions where low soil fertility is an issue (Sheahan, 2012). Tomato (Lycopersicon esculentum, mill) is one of the most important vegetable crops in Egypt and world. Egypt ranks as one of the largest tomato producers in the world. The total cultivated area of tomatoes in Egypt was about 490,260 acres in 2012 (FAO, 2012). Sweet potato, Ipomoea batatas (L.) is an important starchy food crop, especially in developing countries, where it ranks third in value of

production and fifth in calorie contribution to human diets, and constitutes one of the seven most important crops on a worldwide basis (Jones 1970, Chalfant, *et al.* 1990, Jansson and Raman 1991 and FAO 2015).

Unfortunately, during its several growth phases, the aforementioned crops are attacked by a variety of insect pests. Injurious piercing-sucking insect pests seriously impair yield quality and quantity (Jackai, 1995; Ward *et al.*, 2002 and Hassan 2013). They inflict harm directly by suckling on plant juice, or indirectly by acting as viral vectors.

Chemical insecticides have negative consequences on the environment, domestic animals, people, and biological control agents (Schmutterer, 1990). Thus, biological control is still a crucial part in managing insect pests. In the case of vegetables, they are advised as they are almost certainly used as fresh foods. The Ministry of Agriculture has been working to reduce the amount of insecticides used in integrated pest management programs during the past few years. To maintain the natural balance, it is imported to conserve the natural enemies.

According to studies by Helal *et al.* (1996), Abd El-Kareim *et al.* (2011), Salman *et al.* (2014), Khuhro *et al.* (2012), and Al-Deghair *et al.* (2014), the primary cause of death for piercing-sucking insect pests (e.g., aphids, white fly, and leafhopper) is the aphidophagous predators. Predatory coccinellids are important bio-control agents because they feed on a range of phytophagous insect pests, including aphids, scale insects, mealy bugs, mites, white flies, thrips, etc., (Omkar and Pervez, 2002). Coccinellids are considered to be among the most important biological control agents (Ceryngier and Hodek, 1996). The mirid bug (family Miridae) is one of whiteflies main natural enemies. The omnivorous species in this family are significant natural adversaries of several pests in greenhouse crops and solanaceous fields, such as whiteflies (Albajes and Alomar, 1999). Orius spp. (Heteroptera: Anthocoridae) are native predators in Europe (van Lenteren, 1997). They can consume a variety of softbodied arthropods, including aphids (Reitz et al., 2006). Also they can act as biocontrol particularly in greenhouse agents, environments (Rajabpour et al., 2011; Salehi et al., 2016). One of the largest families of order Diptera is Syrphidae, it is most famous for its amazing imitation of wasps and bees. The family Syrphidae has a wide range of feeding preferences; however the Syrphidae subfamily is a significant predator of aphids and other Homoptera as pests (Chambers, 1988). After bees, flies are typically the second most significant visitors to flowers (Larson et al., 2001). *Aphidoletes* aphidimyza (Rondani) (Diptera: Cecidomviidae) has been used as an biological effective control agent in greenhouses for more than 40 years. (Markkula 1963 and Harris 1973). The common green lacewing, Chrysoperla carnea (Stephens) (Neuroptera: Chrysopidae), is one of the most frequent arthropod predators. It feeds on a range of soft-bodied insects, including aphids, scale insects, whiteflies, mites, and the eggs and neonates of lepidopteron insects (McEwen et al. 2001).

The host plant has a significant impact on populations of piercing-sucking pests and their predators. The natural enemies displayed variations in their search characteristics in response to the species of host plant (Abd El-Kareim, 2002). In order to implement IPM programs, the host plant must be taken into account (Marouf, 2007 and Abdel-kareim *et al.*, 2011).

Therefore, the following topics were the focus of the current research:

1. Studying the seasonal abundance of the main predatory insect species and their prey (*Aphis* spp., *Bemisia tabaci*, *Thrips tabaci*, *Empoasca* sp., *Phenacoccus solenopsis* and *Nezara*

viridula) on cucumber, cowpea, tomato and sweet potato crops.

2. Assessing the relationship between prey, host plants, and the seasonal activity of related predators.

MATERIALS AND METHODS

Field experiments:

A private farm located in Kafr-Saad area, Damietta Governorate, Egypt was used to study the relationship between the predatory insects; (Coccinella undecimpunctata, С. septempunctata, Cryptolemus montiziri and Chilocorus nigritus; Orius spp., Macrolophus sp., Aphidoletes aphidimyza, Syrphus sp. and Chrysoperla carnea) and piercing-sucking insect pests (Aphis spp., Bemisia tabaci, Thrips tabaci, Empoasca sp., Phenacoccus solenopsis and Nezara viridula) on different vegetable crops; i.e. cucumber (Cucumis sativus crop, L.: Cucurbitaceae); cowpea crop, (Vigna unguiculata (L.) Walp.: Fabaceae); tomato crop, (Lycopersicon esculentum Mill.: Solanaceae) and sweet potato crop, (Ipomoea batatas L. Lam: Convolvulaceae). To estimate the piercingsucking insects seasonal abundance and predatory insects on the four vegetable plants; cucumber, cowpea, tomato and sweet potato, an area of about 1050 m2 was prepared, as recommended, and divided into four equal plots (each of 262.5 m²), each plot was planted with one of the aforementioned vegetable crops. All vegetable crops were sown on the 27th of April 2022. All regular agricultural practices on the four crops were typically without carried out using pesticides. Throughout the whole production period, all recommended agricultural practices were adhered to, with the exception of using pesticides.

Sampling techniques:

After two weeks of planting date and continuing until harvest, weekly random samples of plants were taken from each replicate. After being placed in polyethylene bags in the field, leaves were collected and brought into the laboratory for analysis. The collected leaves were examined under a stereoscopic microscope. Insects were counted individually on both sides of the leaves. All of the previously stated insect species were counted and their numbers were noted. Predators were noted to have been present on the samples that were gathered. Predatory insects were counted on 25 plants in the field, in addition to leaf samples

Statistical analyses:

Utilizing SPSS Statistics (2020), simple correlation and multiple partial regression analyses were performed between prey densities (i.e., the weekly average of each prey density) and the seasonal abundance of insect predators.

RESULTS AND DISCUSSION

Relative abundance of vegetable crop insect pests:

The obtained results presented in Table (1) show that the vegetable crops i.e. cucumber, cowpea, tomato, and sweet potato were attacking by many insect species especially piercing-sucking insects. The insect species that were recorded belong to two insect orders: Thysanoptera, (Thrips tabaci) and Hemiptera, (Aphis gossypii and Myzus persicae); the tomato whitefly, Bemisia tabaci (Alevrodidae); leafhoppers Empoasca spp. (Cicadellidae); the green stink bug Nezara viridula (L.) (Pentatomidae) and the cotton mealybug, Phenacococcus solenopsis Tinslev (Pseudococcidae).

Only five insect species, *A. gossypii, B. tabaci, Empoasca* spp., *P. solenopsis,* and *N. viridula* were recorded on each of the studied vegetable crops. The most attractive host plant was cucumber (attracted seven insect species) followed by cowpea and tomato (attracted six insect species) whereas sweet potato attracted five insect species.

The most abundant insect species on the studied vegetable crops were *Aphis* spp. recorded the highest number and ratio and presented by 242.4 individuals (51%). followed by the whitefly, *B. tabaci* presented by 161.4 individuals (34.1%). whereas, *N. viridula* was the smallest and represented by 3.2 individuals (0.7%) during the study season.

The relation between predatory insects and the tested host plants:

Total number and relative abundance of predatory insects:

The data in Table (2) and Fig. (1) show that there are nine predator species recorded on the four tested vegetable crops belonging to four orders: the first order is Copleoptrea (*Coccinella undecimpunctata, C. septempunctata, Cryptolemus montiziri* and *Chilocorus nigritus*), the second order is Heteroptera (*Orius* spp. and *Macrolophus* sp.), the third orders is Diptera (*Aphidoletes aphidimyza* and *Metasyrphus* sp.), Nuroptera was found to be the fourth order witch was represented by *Chrysoperla carnea* Table (2).

The data shown in Table (2) revealed that the most abundant predators on the tested host plants were the coccinellid followed by Macroliphus sp. and Orius spp. while, A. *aphidimyza* followed coccinillid only on cucumber plants while on the other three crops it came in fifth category. Syrphus sp. came in the last category of predaceous insects on all of the aforementioned crops. On the other hand, 26.8, 29.7, 29.9, and 26.7% of all predatory insects were coccinellid predators on cucumber, cowpea, tomato and sweet potato respectively. While, crops, Syrphus sp. recorded the smaller number and represented by 5.4, 9.2, 11.2 and 10.6% of the total numbers of predators in cucumber, cowpea, tomato and sweet potato crops, respectively, during the study season.

All predatory insects showed a preference for cucumber, followed by cowpea, while sweet potato plants showed a less preference for all predatory insects. This may be explained by the lower population density of insects that these predators feed on sweet potato plants compared to cucumber plants. The coccinellid predators preferred cucumber (1209 predators) in comparing with the other three host plants, cowpea (225 predators), tomato (123 predators) and sweet potato (78 predators), respectively (Table,1). On the other hand, Heteropteran predators, Macroliphus sp. and Orius sp. showed the same trend, Macroliphus sp. represented by 838, 146, 72 and 64 predators while, Orius sp. represented by 575, 119, 62 and 40 predators on cucumber, cowpea, tomato and sweet potato crops, respectively. Dipteran predators, showed the same trend, Syrphus sp. represented by 243, 70, 46 and 31 predators while, A. aphidimyza represented by 1056, 53, 32 and 21 predators on cucumber, cowpea, tomato and sweet potato crops, respectively. Nuropteran predator showed the same trend, Chy. Carnea population on cucumber, cowpea, tomato and sweet potato plants were 583, 144, 76 and 58 predators, respectively.

Seasonal abundance of predator species on different vegetable crops:

The Coleopteran predators:

Family Coccinellidae:

The data represented in Figure (2), showed that early than the other predators on May 16, the coccinellid predators began to visit cowpea and cucumber plants and recorded three peaks of infestation. The highest peak on cucumber (163 individual) were recorded in the 1st of Aug., while on cowpea and tomato the peak (28 and 16 individual) recorded on the 25th of Jul., The coccinellid predators appear later on June 6th on sweet potato plants, its peak recorded on July 18th with 11 indivi./ plant sample.

Nuropteran predators:

Family Chrysopidae:

As shown in Figure (2), the green lacewings *Chy. carnea* population recorded the highest numbers on the 13th of Jun. 2022 with 82 indivi./ plant sample on cucumber plants. While, on cowpea, tomato and sweet potato plants the highest number of individuals were recorded on Jul. 11 with 15, 9 and 7 indivi./ plant sample.

Dipteran predators:

A. Family: Cecidomyiidae:

The data represented in Figure (2), the aphidophagous gall midge, *Aphidoletes aphidimyza* (Rondani) started to visit the four vegetable crops after about nine weeks on 18th of Jul. and recorded the highest number on 1sth of Aug. with 101 indiv. /sample on cucumber, while on cowpea, tomato and sweet potato plants the highest numbers recorded on Aug. 8 with 9, 5 and 4 individuals/ plant sample.

B. Family: Syrphidae:

The data represented in Figure (2) the syrphid predator, *Syrphus* sp. started to visit the four vegetable crops after about three weeks on 6th of Jun. and recorded the highest number on 8th of Aug. with 38, 9, 6 and 4 indivi./ plant sample on cucumber, cowpea, tomato and sweet potato plants respectively.

Heteropteran predators:

A. Family: Miridae:

The data represented in Figure (2), the mirid bug, *Macroliphus* sp. appeared on cucumber and cowpea crops on 23rd of May and on 30th of May on tomato and sweet potato while, exhibited a distinct peak on cucumber, cowpea and tomato plants (205, 24 and 13 individual/sample) recorded on the 4th of Jul. While on sweet potato plants the peak recorded on Jun. 27 with by 9 indivi./ plant sample.

B. Family: Anthocoridae:

The data represented in Figure (2), the anthocorids pirate, bugs *Orius* spp. appeared on cucumber and cowpea on 30th of May and

on 6th of Jul. on tomato and sweet potato. One peak recorded on cucumber plants (97 individual/ sample) at the 27th of Jun. While, on cowpea, tomato and sweet potato plants the peak recorded on 18th of Jul. with 21, 12 and 7 individual/sample.

The relation between predatory insects and their prey.

Seasonal abundance of predators in response to prey density:

The data represented in Figures (3, 4, 5 and 6) show the population changes of each predator species in response to the densities of their prey on each host plant.

The interaction between seasonal activity of predators, preys and host plants.

Multiregression analysis was used to assess the relationship between the different prey densities and the seasonal activity of associated predatory insects on the examined host plants. The multi-regressions calculated values representing the common effect of average preys densities (i.e. thrips, whitefly, aphids, green stink bug, cotton mealybug and leafhoppers) on the population density of each predator on the four vegetable crops; cowpea, cucumber, sweet potato and tomato plants are displayed in Tables (3, 4, 5 and 6) which indicated to the following:

On cucumber crop:

Data presented in Table (3), show the predaceous relation between insect populations and tested prey on cucumber plants. In the case of coccinellid predators, this relation showed significant correlation with Aphis spp., T. tabaci, Empoasca spp. and N. viridula populations, the correlation coefficient value (r) = 0.524, -0.496, -0.588 and 0.632 respectively, while insignificant correlation with B. tabaci and P. solenopsis populations (r) =0.284 and 0.397. While, this relation in the case of Macroliphus sp. and Orius sp. were insignificant with all prev except N. viridula populations was highly significant and positive, the correlation coefficient value (r) =0.612 and 0.771 respectively. On the other hand, Syrphus sp. this relation were highly significant with all prey except Empoasca spp. populations was insignificant and negative, the correlation coefficient value (r) = -0.373. Whereas, A. aphidimyza this relation were significant with B. tabaci and T. tabaci while, highly significant correlation with *P. solenopsis*, the correlation coefficient value (r) = 0.533, 0.445 and 0.772 respectively. Finally, C. carnea this relation were non-significant with each

prey except *N. viridula* populations was significant and positive, the correlation coefficient value (r) =0.580.

As shown in (Table 3) multi-regression analysis revealed that the common effect of the prey population size (i.e. aphids, whitefly, thrips, leafhoppers, cotton mealybug and green stink bug) exhibited strong effect on the predatory insect populations.

The strongest effect shown in the case of *Syrphus* sp. (91.5%) and the weakest effect shown in the case of *C. carnea* predators (6.8%) of the overall population changes were caused by compound effect of the tested preys. Multi-regression analysis showed that the common effect of the population size of the aforementioned prey on the coccinellid, *Macroliphus* sp., *Orius* sp. and *A. aphidimyza* populations were 55.2, 50.0, 64.1 and 73.0% of the overall population changes were caused by the tested prey compound effect.

On cowpea crop:

Data presented in Table (4), show the relation between predaceous insect populations and tested prey on cowpea plants. In the case of coccinellid predators, this relation showed highly significant and positive correlation with P. solenopsis and N. viridula populations, the correlation coefficient value (r) = 0.938 and 0.930 respectively, while insignificant correlation with Aphis spp., B. tabaci, T. tabaci and Empoasca spp. populations. While, this relation in the case of Macroliphus sp. predator were significantly positive with Aphis spp. and P. solenopsis and highly significant with Empoasca spp. and N. viridula, the significantly coefficient value (r) = 0.581, 0.532, 0.622 and 0.790 respectively. This relation was insignificant with *B. tabaci* and *T.* tabaci. This relations in the case of Orius sp. predators with P. solenopsis and N. viridula insects were highly significant and positive, the correlation coefficient value (r) = 0.698 and 0.869 respectively, non-significant correlation were shown with the other insect species. On the other hand, the predatory insect, Syrphus sp. this relation was highly significant with *T*. tabaci and P. solenopsis and significant with N. viridula the correlation coefficient value (r) = -0.629, 0.702 and 0.498, while, it this relation was insignificant with Aphis spp., B. tabaci and Empoasca spp. Whereas, A. aphidimyza this relation was significant with T. tabaci, Empoasca spp and *P. solenopsis*, the correlation coefficient value (r) = -0.603, -0.566 and 0.461 respectively, while, non-significant correlation with Aphis spp, B. tabaci and N. viridula. The correlation

coefficient value between *C. carnea* and each of *Aphis* spp. and *N. viridula* were highly significant and significantly positive with *B. tabaci*, the correlation coefficient value (r) = 0.644, 0.695 and 0.488 respectively, but it was insignificant with *T. tabaci*, *Empoasca* spp. and *P. solenopsis*.

As shown in (Table 4) multi-regression analysis revealed that the common effect of the prey population size (i.e. aphids, whitefly, thrips, leafhoppers, cotton mealybug and green stink bug) exhibited strong effect on the predatory insect populations.

The strongest effect shown in the case of coccinellid predators (91.3%) and the weakest effect shown in the case of *A. aphidimyza* predators (32.5%) of the overall population changes were caused by compound effect of the tested preys. Multi-regression analysis showed that the common effect of the population size of the aforementioned prey on the *Macroliphus* sp., *Orius* sp., *Syrphus* sp. and *C. carnea* populations were 79.3, 76.1, 54.6 and 72.6% of the overall population changes were caused by the tested prey compound effect.

On tomato crop:

Data presented in Table (5), show the predaceous relation between insect populations and tested prey on tomato plants. In the case of coccinellid predators, this relation showed highly significant correlation with *N. viridula* and significantly positive with Aphis spp. populations, the correlation coefficient value (r) = 0.932 and 0.512respectively, while insignificant correlation with B. tabaci, T. tabaci, Empoasca spp. and P. solenopsis populations. While, this relation in the case of *Macroliphus* sp. predator was highly significant and positive with N. viridula and significantly negative with P. solenopsis, the correlation coefficient value (r) = 0.741 and -0.537 respectively. This relation was insignificant with Aphis spp., B. tabaci, T. tabaci and *Empoasca* spp. This relations in the case of Orius sp. predators with N. viridula insects were highly significant and positive, the correlation coefficient value (r) = 0.891, nonsignificant correlation were shown with the other insect species. On the other hand, the predatory insect, Syrphus sp. this relation was highly significant and positive with *Aphis* spp. and N. viridula and significant with B. tabaci and T. tabaci, the correlation coefficient value (r) = 0.746, 0.677, 0.514 and -0.587, while, it this relation was insignificant with *Empoasca* spp. and P. solenopsis. Whereas, A. aphidimyza this relation was highly significant and positive

with *Aphis* spp. and significant with *B. tabaci*, *T. tabaci* and *P. solenopsis*, the correlation coefficient value (r) = -0.632, 0.533, -0.586 and 0.543 respectively, while, non-significant correlation with *Empoasca* spp. and *N. viridula*. The correlation coefficient value between *C. carnea* and *N. viridula* was highly significant and significantly positive with *Aphis* spp. and *Empoasca* spp., the correlation coefficient value (r) = 0.755, 0.584 and 0.441 respectively, but it was non-significant with *B. tabaci*, *T. tabaci* and *P. solenopsis*.

As shown in (Table 5) multi-regression analysis revealed that the common effect of the prey population size (i.e. aphids, whitefly, thrips, leafhoppers, cotton mealybug and green stink bug) exhibited strong effect on the predatory insect populations.

The strongest effect shown in the case of *Syrphus* sp. followed by coccinellid and *Orius* sp. predators with 86.7, 85.4 and 82.3% of the overall population changes were caused by compound effect of the tested preys and the weakest effect shown in the case of *A. aphidimyza* predators (51.2%) of the overall population changes were caused by compound effect of the tested preys. While, *Macroliphus* sp. and *C. carnea*, multi-regression analysis showed that the common effect of the population size of the aforementioned prey were 56.7 and 53.3% of the overall population changes were caused by the tested prey compound effect.

On sweet potato crop:

Data presented in Table (5), show the between predaceous relation insect populations and tested prey on sweet potato plants. In the case of coccinellid predators, this relation showed highly significant correlation with *N. viridula* and significantly positive with spp. populations, the correlation Aphis coefficient value (r) = 0.934 and 0.508respectively, while insignificant correlation with B. tabaci, Empoasca spp. and P. solenopsis populations. While, this relation in the case of Macroliphus sp. predator was highly significant and positive with N. viridula and significant with B. tabaci, Empoasca spp and P. solenopsis, the correlation coefficient value (r) = 0.838, 0.468, 0.489 and -0.517 respectively. This relation was insignificant with *Aphis* spp. This relations in the case of Orius sp. predators with N. viridula insects were highly significant and positive, the correlation coefficient value (r) =0.942, non-significant correlation were shown with the other insect species, Aphis spp., B. tabaci, Empoasca spp and P. solenopsis. On the

other hand, the predatory insect, Syrphus sp. this relation was highly significant and positive with Aphis spp. and N. viridula, the correlation coefficient value (r) = 0.729 and 0.621, while, it this relation was insignificant with B. tabaci, Empoasca spp. and P. solenopsis. Whereas, A. aphidimyza this relation was significantly positive with Aphis spp. and P. solenopsis, the correlation coefficient value (r) = 0.600 and 0.527 respectively, while, nonsignificant correlation with *B. tabaci, Empoasca* spp. and N. viridula. The correlation coefficient value between C. carnea and N. viridula was highly significant and significantly positive with Aphis spp., B. tabaci and Empoasca spp., the correlation coefficient value (r) = 0.791, 0.574, 0.478 and 0.489 respectively, but it was non-significant and negative with, *P. solenopsis*.

As shown in (Table 6) multi-regression analysis revealed that the common effect of the prey population size (i.e. aphids, whitefly, thrips, leafhoppers, cotton mealybug and green stink bug) exhibited strong effect on the predatory insect populations.

The strongest effect shown in the case of *Orius* sp. followed by coccinellid and *Macroliphus* sp. predators with 94.8, 89.0 and 81.6% of the overall population changes were caused by compound effect of the tested preys and the weakest effect shown in the case of *A. aphidimyza* predators (53.0%) of the overall population changes were caused by compound effect of the tested preys. While, *Syrphus* sp.and *C. carnea*, multi-regression analysis showed that the common effect of the population size of the aforementioned prey were 69.6 and 64.0% of the overall population changes were caused by the tested prey compound effect.

DISCUSSION

Attractiveness of different host plants to predatory insects

Surveying the predatory insects that associated with the summer planting season of the four vegetable crops (tomato, cucumber, cowpea and sweet potato) assured that the most abundant insect predators recorded were the coccinellid, (*Cryptolemus montiziri, C. septempunctata, C. undecimpunctata* and *Chilocorus nigritus*), Orius spp., Macrolophus sp., A. aphidimyza, C. carnea and Syrphus sp.

As previously noted by other authors, these predators were identified as important natural enemies of vegetable plants sweet potato, cucumber, cowpea, and tomato crops in Egypt, (Abdel-Gawaad *et al.*, 1990; Amro 2004; Ali *et* *al.*, 2013 and Gameel 2013 El-Fakharany, *et al.* 2017).

According to the current study, predaceous insects' seasonal abundance showed variations in their response to different host plants. The collected predators greatly preferred cucumber over cowpea, tomato and sweet potato. Also, the predacious insects; C. motrouzari, Chilocorus bipustulatus and Rodolia cardinalis showed differences in its response to different host plants (Cardosa, 1990; Heidari et al., 1999 and Abdel-Mageed, 2005). The average number of C. undecimpunctata and Scymnus significant syriacus showed differences between tomato cultivar Yassin et al. (2014).

According to El-Baradey (2012), differences in the type of Kairomone that produce by different plant species may cause in variation of predator response to different host plants. Abd El-Kareim (2002); Abdel-Kareim et al., (2011) and Marouf (2011) came to the same result, the main factor in insect attraction to host plant is emission of auditory stimuli. According to Luna and Jepson (2001), variations in hoverflies and coccinellid beetles response to the host plants under investigation could be caused by either chemical or physical stimulation. A combination of substances, including both the volatiles from the prey and the plants in the habitat, are involved for some predatory insects (Hagen, 1986). Satti and Mahgoub (2018) recorded four predators; the syrphid fly, Xanthogramma aegyptium, C. undecimpunctata, C. carnea, and H. variegate associated with T. tabaci on tomato, rocket, and onion plants. Even though hundreds of predators have been known to attack B. tabaci, the most frequent ones are as follows: lacewings (C. carnea and C. pallens), bugs (Orius sp., Macrolophus caliginosus, and Nesidiocoris tenuis), in addition to mites (Amblyseius swirskii and Euseius ovalis) Al-Zyoud (2014).

Interaction between predators activity, preys and host plants.

The geographical distributions of organisms within biological communities are shaped significantly by predator-prey interactions (Williams and Flaxman, 2012). Significant host-prey interactions were shown by Jalali and Michaud (2012) to be present for all aspects of development, including juvenile survival, developmental time, adult mass at emergence, and associated predator reproduction.

In this study, tested predatory insects, i.e. *Orius* species, Coccinellids, *Macrolophus*

species, Aphidimyza aphidimyza, Syrphus species, and C. carnea showed differences in their response to the insect prey population on the different host plants, especially with populations of the leafhopper. When the coccinillid predator, C. undecimpunctata fed on the cotton aphid, A. gossypii showed higher rates of searching than when fed on the pomegranate aphid, Aphis punicae (Al-Deghair et al., 2014). Also, when the larval stage of the aphid lion, Chrysoperla carnea fed on the different aphid species (the English grain aphid, Sitobion avenae, the cotton aphid, A. gossypii, oleander aphid, Aphis nerii and corn aphid, Rhopalosiphum maidis) the total duration of the immature stages differed significantly (El-Serafi et al., 2000). According to Giles et al. (2002), the pea aphid Acyrthosiphon pisum fed on Alfalfa (Medicago sativa) it was suitable prey for the seven-spot ladybird (*C. septempunctata*) survival, developmental time, and size of the adult in comparing with the same species reared on the broad bean (Vicia faba L.) The developmental rate was significantly faster when the coccinillid predator, the seven-spot ladybird (C. septempunctata) was reared on A. pisum in comparing with the same species reared on R. maidis (Obrycki and Orr, 1990). According to Cottrell and Tillman (2017) four species of lady beetles (Coleoptera: Coccinellidae) exhibit limited predation on N. viridula eggs and nymphs.

REFERENCES

- Abd El-Kareim, A.I. 2002: The potential of some natural enemies as bioagents against certain diaspidid species. J. Union Arab Biol. Cairo, 17 (A): Zoology, 51-63.
- Abd El-Kareim, A.I., El-Naggar, M.E., Amal, E. Marouf 2011: Survey of predaceous insects associated with four medicinal plants. J. plant prot. And Pathology, Mansoura Univ., Vol. 2(6): 623-636.
- Abdel-Gawaad, A.A., El-Sayed, A.M., Shalaby, F.F., and Abo El-Ghar, M.R. 1990: Natural enemies of *Bemisia tabaci* and their role in suppressing the population density of the pest. Agric. Res. Rev. 68(1): 185-195.
- Abdel-Mageed, Sanaa, A.M. 2005: Influence of certain natural enemies on some mealybug populations. M. Sc. Thesis, Fac. Agric., Mansoura Univ., Mansoura, Egypt, P. 154.
- Albajes, R., Alomar, Ò. 1999: Current and potential use of polyphagous predators. In Integrated pest and disease management in greenhouse crops. Dordrecht: Springer Netherlands. (pp. 265-275).
- Al-Deghair, M.A., Abdel-Baky, N.F., Fouly, A.H., Ghanim, N.M. 2014: Foraging behavior of Two

coccinellid species (Coleoptera: Coccinellidae) fed on aphids. J. Agric. and Urban Entomol. 30(1):12-24.

- Ali, SH.A., Saleh, A.A., Mohamed, Nadia, E. 2013: *Aphis craccivora* Koch. and predators on faba bean and cowpea in newly reclaimed areas in Egypt. Egyptian Journal of Agricultural Research, 91(4), 1423-1438.
- Al-Zyoud, F.A. 2014: The most common predators of *Bemisia tabaci* (Genn.): Biology, predation, preferences, releases, alternative food resources, combined use, current efforts and future perspectives. Journal of Biological Control, 28(1), 1-16.
- Amro, M.A.M. 2004: Incidence of certain arthropod pests and predators inhabiting cowpea, with special reference to the varietal resistance of selected cultivars to *Bemisia tabaci* (GEN.) and *Tetranychus urticae* KOCH. Ass. Univ. Bull. Environ. Res., 7 (1).
- Belane, A.K., Dakora, F.D. 2009: Measurement of N2 fixation in 30 cowpea (*Vigna unguiculata* L. Walp.) genotypes under field conditions in Ghana, using the15N natural abundance technique. Symbiosis 48 (1-3): 47-56.
- Cardosa, A. 1990: Preliminary study of the coccinellids found on citrus in Portugal. Boletin de Sanidad Vegetal, Plagas, 16(1): 105-111.
- Ceryngier, P., Hodek, I. 1996: Enemies of the Coccinellidae. In: Hodek, I., Honk, A. (Eds.), Ecology of Coccinellidae. Kluwer Academic Publishers, Dordrecht, pp. 319-350.
- Chalfant, R.B., Jansson, R.K., Seal, D.R., Schalk, J.M. 1990: Ecology and management of sweet potato insects. Annual review of entomology, 35(1), 157-180.
- Chambers, R.G. 1988: Applied production analysis: a dual approach. Cambridge University Press.
- Cottrell, T.E., Tillman, P.G. 2017: Four species of lady beetles (Coleoptera: Coccinellidae) exhibit limited predation on *Nezara viridula* (Hemiptera: Pentatomidae) eggs and nymphs. *Biological Control*, 114, 73-78.
- El-Baradey, Wafaa, M.M. 2012: Eco-physiological studies on some scale insects. PhD. Sc. Thesis, Fac. Agric., Mansoura Univ., Mansoura, Egypt, P. 155.
- El-Fakharany, S.K., Hegazy, F.H., Samy, M.A.E.M. 2017: Survey and Population Fluctuations of Arthropod Pests and Predators in Sweet Potato at Nile Delta, Egypt. Egyptian Academic Journal of Biological Sciences. A, Entomology, 10 (7), 277-285.
- El-Serafi, H.A.K., Abdel-Salam, A.H., Abdel-Baky, N.F. 2000: Effect of four aphid species on certain biological characteristics and life table parameters of *Chrysoperla carnea* Stephens and

Chrysopa septempunctata Wesmael (Neuroptera: Chrysopidae) under laboratory conditions .Pakistan J. Biolog. Sci.

- FAO, 2012: "Statistical Yearbook". Food and Agriculture Organization (FAO), Rome, Italy.
- FAO, 2015: Food and Agriculture Organization of the United Nations (FAO), Food and Agriculture Organization Statistical Databases (FAOSTAT), 2015, http://faostat3.fao.org/browse/O/OC/E.
- Gameel, S.M.M. 2013: Species composition of piercing-sucking arthropod pests and associated natural enemies inhabiting cucurbit fields at the new valley in Egypt. Egypt. Acad. J. Biolog. Sci., 6(2): 73 – 79.
- Giles, K.L., Madden, R.D., Stockland, R., Payton, M.E., Dillwith, J.W. 2002: Host plants affect predator fitness via the nutritional value of herbivore prey: Investigation of a plant-aphidladybeetle system. J. Biol. Control., 47: 1-21.
- Hagen, K.S. 1986: Ecosystem analysis: plant cultivars (HRP), entomophagous species and food supplements. In: Boethal, D.J., Eikenbary, R.D. (Eds.), Interactions of Plant Resistance and Parasitoids and Predators of Insects. John Wiley & Sons, New York, pp. 151–197.
- Harris, K.M. 1973: Aphidophagous Cecidomyiidae (Diptera): taxonomy, biology and assessments of field populations. B. Entomol. Res. 63:305-325.
- Hassan, S. 2013: Effect of variety and intercropping on two major cowpea [*Vigna unguiculata* (L.) Walp] field pests in Mubi, Adamawa State, Nigeria. Int. J. Agric. Res. Dev. 1(5): 108-109.
- Heidari, M., Hodgson, C., Porcelli, F. 1999: Influence of host-plant physical defenses on the searching behavior and efficacy of two coccinellid predators of the obscure mealybug, *Pseudococcus viburni* (Signoret). Entomolologica, 33: 397-402.
- Helal, A.H., Salem, R.M., El- khouly, A.S., Metwally, M.M., ElMezaien, A.B. 1996: Population dynamic of *A. craccivora* Koch and *Empoasca* spp. on Faba bean in relation to associated predators and some climatic factors. Egypt. J. Agric. Res. 75(2): 461-471.
- IBM Corp. 2020: IBM SPSS Statistics for Windows (Version 27.0) [Computer software]. IBM Corp.
- Jackai, L.E.N. 1995: The legume pod borer *Maruca testulalis* and its principal host plant, *Vigna unguiculata* (L.) walp. Use of selective insecticide sprays as an aid in the identification of useful levels of resistance. Crop Prot. 14: 4, 299-306.
- Jalali, M.A., Michaud, J.P. 2012: Aphid-plant interactions affect the suitability of *Myzus* spp. as prey for the two spot ladybird, *Adalia*

bipunctata (Coleoptera: Coccinellidae). Eur. J. Entomol., 109: 345-352.

- Jansson, R.K., Raman, K.V. 1991: Sweet Potato Pest Management: A Global Perspective. Westview Press, Boulder, CO.
- Jones, A. 1970: The sweet potato-today and tomorrow, pp. 3-6. In D. L. Plucknett (ed.). Proceedings of the 2nd symposium for the International Society of Tropical Root Crops. Vol. 1., University of Hawaii, Honolulu.
- Khuhro, N.H., Chen, H., Zhang, Y., Zhang, L., Wang, M. 2012: Effect of different prey species on the life history parameters of *Chrysoperla sinica* (Neuroptera: Chrysopidae). Eur. J. Entomol., 109: 175-180.
- Larson, B.M.H., Kevan, P.G., Inouye, D.W. 2001: Flies and flowers: taxonomic diversity of anthophiles and pollinators. The Canadian Entomologist, 133(4), 439-465.
- Luna, J., Jepson, P. 2001: Organic farming research project report submitted to: organic farming research foundation. Project title: Enhancement of biological Control with Insectary Planting. Santa Cruz.
- Markkula, M. 1963: Studies on the pea aphid, *Acyrthosiphon pisum* Harris (Hom, Aphididae) with special reference to the difference in biology of the green and red forms. Ann. Agr. Fenn. 2:1-30.
- Marouf, Amal, H. 2007: Studies on insect pests and their natural enemies associated with marjoram and chamomile plants. M. Sc. Thesis, Fac. Agric., Mansoura Univ., Mansoura, Egypt, P. 150.
- Marouf, Amal, H. 2011: Influence of some medicinal plants as a habitat management tool to enhance the role of certain natural enemies. Phd. Sc. Thesis, Fac. Agric., Mansoura Univ., Mansoura, Egypt, P. 164.
- McEwen, P.K., New, T.R.R., Whittington, A. 2001: Lacewings in the crop management. Cambridge University Press. 546p, 14, 1806-1812.
- Obrycki, J.J., Orr, C.J. 1990: Sutability of three prey species for neaecticpo pulation of *Coccinella septempunctata*, *Hippodamia variegata* and *Propylea quatuordecimpunctata* (Coleoptera: Coccinellidae). J. Econ. Entomol. 83(4): 1292-1297.
- OEPP/EPPO, 27: 15-27.
- Omkar, Pervez, A. 2002: New record of coccinellids from Uttar Pradesh .III. Journal of Advanced Zoology. 23(1): 63-65.
- Rajabpour, A., Seraj, A.A., Allahyari, H., Shishehbor, P. 2011: Evaluation of *Orius laevigatus* Fiber (Heteroptera: Anthocoridae) for biological control of *Thrips tabaci* Lindeman

(Thysanoptera: Thripidae) on greenhouse cucumber in South of Iran. Asia Journal of Biological Science, 4: 457-467.

- Reitz, S.R., Funderburk, J.E., Scot, M.W. 2006: Differential predation by the generalist predator *Orius insidiosus* on congeneric species of thrips that vary in size and behavior. Entomologia Experimentalis et Applicata, 119: 179-188.
- Salehi, Z., Yarahmadi, F., Rasekh, A., Sohani, N.Z. 2016: Functional responses of Orius albidipennis Reuter (Hemiptera, Anthocoridae) to Tuta absoluta Meyrick (Lepidoptera, Gelechiidae) on two tomato cultivars with different leaf morphological characteristics. Entomologia Generalis, 36: 127-136.
- Salman, A.M.A., El-Harery, M.A., El-Solimany, E.A. 2014: Effects of population densities of *Aphis cracivora* Koch. on predatory efficiency of *Coccinella septempunctat* L., *Coccinella undecimpunctata* L. and *Chrysoperla carnea* Stephens larvae under laboratory conditions. Middle East J. Agric. Res. 3(1): 116-122.
- Satti, A.A., Mahgoub, H.A.H. 2018: Population abundance of *Thrips tabaci* Lindeman and its associated predators on some crops at Shendi, River Nile State, Sudan.International Journal of Scientific Progress and Research (IJSPR), 54, (1):37-45.
- Schmutterer, H. 1990: Properties and potential of natural pesticides from the neem tree, *Azadirichta indica*. Ann. Rev Entomol. 35: 271-289.
- Sheahan, C.M. 2012: Plant guide for cowpea (*Vigna unguiculata* L. Walp) USDA- Natural Resources Conservation Service, Cap May Plant Materials Center, Cap May, Nj.
- Van Lenteren, J.C. 1997: Benefits and risks of introducing exotic macro-biological control agents into Europe.- Bulletin
- Ward, A., Morse, S., Denholm, I., Mc. Namara, N. 2002: Foliar insect pest management on cowpea (*Vigna ungiculata* (L.) Walpers) in simulated varietal mixtures. Field-Crops-Research. 79: 1, 53-65.
- Wargovich, M.J. 2000: Anticancer properties of fruits and vegetables. HortScience, 35(4), 573-575.
- Williams, A.C., Flaxman, S.M. 2012: Can predators assess the quality of their prey's resource?. J. Animal Behaviour 83: 883-890.
- Yassin, S.A., Arafat, N.F., Baiomy, F.A. 2014: Susceptibility of some tomato varieties to some pests and predators. Egyptian Academic Journal of Biological Sciences. A, Entomology, 7(1), 1-10.

Dumetta Governorate.													
Host plant	Aphis spp.		B. tabaci		T. tabaci		Empoasca spp.		P. solenopsis		N. viridula		Tatal
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	Total
Cucumber	890.1	52.5	631.2	37.2	142.4	8.4	6.4	0.4	24.6	1.5	1.8	0.1	1696.5
Cowpea	47.8	42.8	5.1	4.6	4.4	3.9	47	42.1	4.3	3.8	3.1	2.8	111.7
Tomato	29.7	46.9	5.4	8.5	11	17.4	3.9	6.2	9.9	15.6	3.4	5.4	63.3
Sweet potato	1.9	8.5	3.9	17.5	0.0	0.0	4.7	21.1	7.4	33.2	4.4	19.7	22.3
Mean	242.4	51.2	161.4	34.1	39.5	8.3	15.5	3.3	11.6	2.4	3.2	0.7	473.5

Table 1: Mean number and relative abundance of insect species during 2022 at Kafr-Saad region Damietta Governorate.

Table 2: Total number and relative abundance of predatory insect species on vegetable crops during 2022 at Kafr-Saad region Damietta Governorate.

Orders	Coleo	ptrea	Heteroptera				Diptera				Nuroptera		_
Family	Coccin	ilidae	Miridae		Anthocoridae		Syrphidae		Cecidomyiidae		Chrysopidae		
Host plant	Coccilillid		Macroliphussp. Orius sp.		Syrphus sp. A. aphidimyz		limyza	C. carnea		Total			
	No.	%	No.	%	No.	%	No.	%	No.	%	No.	%	-
Cucumber	1209	26.8	838	18.6	575	12.8	243	5.4	1056	23.4	583	12.9	4504
Cowpea	225	29.7	146	19.3	119	15.7	70	9.2	53	7.0	144	19.0	757
Tomato	123	29.9	72	17.5	62	15.1	46	11.2	32	7.8	76	18.5	411
Sweet potato	78	26.7	64	21.9	40	13.7	31	10.6	21	7.2	58	19.9	292
Mean	408.8	27.4	280.0	18.8	199.0	13.3	97.5	6.5	290.5	19.5	215.3	14.4	1491.0

Table 3: The correlation and regression coefficient between the predators (average number) and each of thrips, cotton mealybug, whitefly, leafhoppers, aphids, and green stink bug population densities on cucumber crop during 2022.

· · · ·		Sin	nple	Multiple partial regression					
Predator	Prey	corre	lation						
		r.	Р.	b.	p.	"F"	Prob>F	E.V.	
	Aphis spp.	0.524	0.021	0.02	0.077	4.7	0.011	55.2%	
	B. tabaci	0.284	0.238	-0.02	0.218				
ر د مانین النا ما	T. tabaci	-0.496	0.031	0.05	0.324				
Coccinina	<i>Empoasca</i> spp.	-0.588	0.008	-4.34	0.147				
	P. solenopsis	0.397	0.093	0.95	0.063				
	N. viridula	0.632	0.004	14.06	0.064				
	Aphis spp.	0.124	0.613	-0.01	0.663	4.00	0.02	50.0%	
	B. tabaci	-0.251	0.300	0.01	0.640				
Macroliphus	T. tabaci	-0.217	0.371	-0.08	0.256				
sp.	<i>Empoasca</i> spp.	-0.137	0.575	5.93	0.205				
1	P. solenopsis	-0.386	0.103	-1.36	0.087				
	N. viridula	0.612	0.005	35.31	0.007				
	Aphis spp.	0.349	0.143	0.01	0.358	6.35	0.003	64.1%	
<i>Orius</i> sp.	B. tabaci	-0.202	0.407	-0.01	0.648				
	T. tabaci	-0.306	0.202	-0.01	0.741				
	<i>Empoasca</i> spp.	-0.331	0.167	1.19	0.561				
	P. solenopsis	-0.279	0.247	-0.31	0.367				
	N. viridula	0.771	0.000	21.05	0.001				
	Aphis spp.	0.837	0.000	0.01	0.000	33.35	0.000	91.5%	
	B. tabaci	0.675	0.002	0.01	0.777				
0 1	T. tabaci	-0.485	0.035	0.03	0.558				
Syrphus sp.	<i>Empoasca</i> spp.	-0.373	0.116	-0.41	0.227				
	P. solenopsis	0.630	0.004	0.15	0.014				
	N. viridula	0.489	0.034	1.17	0.163				
	Aphis spp.	0.040	0.869	-0.01	0.713	9.09	0.001	73.0%	
	B. tabaci	0.533	0.019	-0.01	0.482				
А.	T. tabaci	-0.445	0.056	0.01	0.962				
aphidimyza	<i>Empoasca</i> spp.	-0.413	0.079	-8.34	0.026				
1 5	P. solenopsis	0.772	0.000	1.86	0.005				
	N. viridula	-0.038	0.877	-17.62	0.051				
	Aphis spp.	0.389	0.100	0.01	0.552	1.22	0.362	6.80%	
	B. tabaci	0.118	0.630	0.01	0.754				
6	T. tabaci	-0.299	0.213	0.01	0.924				
C. carnea	Empoasca spp.	-0.311	0.196	-0.42	0.854				
	P. solenopsis	0.030	0.903	-0.15	0.691				
	N. viridula	0.580	0.009	9.19	0.118				

Table 4: The correlation and regression coefficient between the predators (average number) and each of thrips, cotton mealybug, whitefly, leafhoppers, aphids, and green stink bug population densities on cowpea crop during 2022.

		Si	mple							
Produtor	Drow	corr	elation]	Multiple partial regression analysis					
rieuator	riey	an	alysis							
		r.	Р.	b.	p.	"F"	Prob>F	E.V.		
	Aphis spp.	0.297	0.217	-0.014	0.537	32.390	0.000	91.3%		
	B. tabaci	0.277	0.251	-0.062	0.735					
Coordinillid	T. tabaci	-0.359	0.131	-0.017	0.944					
Coccimina	Empoasca spp.	0.122	0.619	-0.003	0.959					
	P. solenopsis	0.938	0.000	0.898	0.060					
	N. viridula	0.930	0.000	1.942	0.088					
	Aphis spp.	0.581	0.009	0.083	0.033	12.470	0.000	79.3%		
	B. tabaci	0.409	0.082	-0.423	0.151					
Macroliphus	T. tabaci	0.025	0.921	-0.153	0.680					
sp.	Empoasca spp.	0.622	0.004	0.123	0.141					
	P. solenopsis	0.532	0.019	-0.414	0.543					
	N. viridula	0.790	0.000	2.687	0.119					
Orius sp.	Aphis spp.	0.233	0.338	-0.025	0.421	10.550	0.000	76.1%		
	B. tabaci	0.255	0.293	-0.309	0.213					
	T. tabaci	-0.198	0.415	-0.072	0.821					
	Empoasca spp.	0.410	0.081	0.059	0.396					
	P. solenopsis	0.698	0.001	-0.177	0.760					
	N. viridula	0.869	0.000	2.775	0.065					
	Aphis spp.	0.198	0.416	0.019	0.343	4.610	0.012	54.6%		
	B. tabaci	-0.104	0.672	-0.022	0.886					
Complete and	T. tabaci	-0.629	0.004	-0.274	0.202					
syrphus sp.	Empoasca spp.	-0.363	0.127	0.011	0.806					
	P. solenopsis	0.702	0.001	0.651	0.104					
	N. viridula	0.498	0.030	-0.653	0.479					
	Aphis spp.	-0.123	0.616	-0.004	0.876	2.440	0.089	32.5%		
	B. tabaci	-0.303	0.207	-0.119	0.578					
А.	T. tabaci	-0.603	0.006	-0.018	0.949					
aphidimyza	Empoasca spp.	-0.566	0.012	-0.059	0.337					
	P. solenopsis	0.461	0.047	0.222	0.666					
	N. viridula	0.222	0.361	0.345	0.781					
	Aphis spp.	0.644	0.003	0.080	0.009	8.95	0.001	72.6%		
	B. tabaci	0.488	0.034	-0.091	0.669					
C. comeso	T. tabaci	-0.187	0.444	0.087	0.755					
C. curneu	Empoasca spp.	0.296	0.219	-0.086	0.167					
	P. solenopsis	0.417	0.075	-1.499	0.011					
	N. viridula	0.695	0.001	3.943	0.007					

Table 5: The correlation and regression coefficient between the predators (average number) and each of thrips, cotton mealybug, whitefly, leafhoppers, aphids, and green stink bug population densities on tomato crop during 2022.

		Simple	correlation	Multiple partial regression analysis					
Predator	Prey	aı	nalysis	IVIU	inple pa	Itial regression analysis			
		r.	Р.	b.	p.	"F"	Prob>F	E.V.	
	Aphis spp.	0.512	0.025	0.009	0.550	18.600	0.000	85.4%	
	B. tabaci	0.306	0.203	-0.002	0.995				
	T. tabaci	-0.356	0.135	0.023	0.698				
Coccinillid	Empoasca spp.	0.312	0.194	-0.107	0.516				
	P. solenopsis	-0.082	0.740	0.071	0.414				
	N. viridula	0.932	0.000	1.870	0.000				
	Aphis spp.	0.154	0.529	-0.022	0.362	4.920	0.009	56.7%	
	B. tabaci	-0.148	0.546	0.066	0.879				
Macroliphus	T. tabaci	0.037	0.882	-0.044	0.650				
sp.	Empoasca spp.	0.427	0.068	-0.147	0.583				
	P. solenopsis	-0.537	0.018	-0.211	0.149				
	N. viridula	0.741	0.000	1.239	0.017				
	Aphis spp.	0.277	0.251	-0.019	0.159	14.990	0.000	82.3%	
	B. tabaci	0.026	0.916	0.028	0.904				
0 ·	T. tabaci	-0.158	0.519	-0.013	0.805				
Orius sp.	Empoasca spp.	0.313	0.193	-0.293	0.059				
	P. solenopsis	-0.369	0.120	-0.099	0.202				
	N. viridula	0.891	0.000	1.514	0.000				
	Aphis spp.	0.746	0.000	0.025	0.001	20.630	0.000	86.7%	
	B. tabaci	0.514	0.024	-0.171	0.110				
C 1	T. tabaci	-0.587	0.008	0.006	0.777				
Syrphus sp.	Empoasca spp.	0.039	0.873	0.024	0.708				
	P. solenopsis	0.347	0.145	0.126	0.002				
	N. viridula	0.677	0.001	0.527	0.000				
	Aphis spp.	0.632	0.004	0.030	0.025	4.150	0.017	51.2%	
	B. tabaci	0.533	0.019	-0.121	0.567				
	T. tabaci	-0.586	0.008	0.005	0.907				
A. aphidimyza	Empoasca spp.	-0.263	0.277	-0.043	0.743				
	P. solenopsis	0.543	0.016	0.126	0.084				
	N. viridula	0.268	0.268	0.242	0.287				
	Aphis spp.	0.584	0.009	0.032	0.096	4.43	0.014	53.3%	
	B. tabaci	0.199	0.413	-0.141	0.656				
6	T. tabaci	-0.257	0.289	-0.035	0.621				
C. carnea	Empoasca spp.	0.441	0.059	0.251	0.216				
	P. solenopsis	-0.152	0.534	0.028	0.788				
	N. viridula	0.755	0.000	0.591	0.096				

Table 6: The correlation and regression coefficient between the predators (average number) and each of thrips, cotton mealybug, whitefly, leafhoppers, aphids, and green stink bug population densities on sweet potato crop during 2022.

		Simple	correlation	Multiple partial regression analysis					
Predator	Prey	ar	nalysis						
	-	r.	Р.	b.	p.	"F"	Prob>F	E.V.	
	Aphis spp.	0.508	0.026	0.025	0.849	30.150	0.000	89.0%	
	B. tabaci	0.298	0.215	-0.590	0.671				
Coccinillid	Empoasca spp.	0.306	0.203	0.449	0.701				
	P. solenopsis	-0.038	0.876	0.072	0.071				
	N. viridula	0.934	0.000	0.978	0.000				
	Aphis spp.	0.356	0.135	0.016	0.927	17.000	0.000	81.6%	
	B. tabaci	0.468	0.043	-4.476	0.027				
Macroliphus sp.	Empoasca spp.	0.489	0.034	3.714	0.0.29				
	P. solenopsis	-0.517	0.023	-0.116	0.033				
	N. viridula	0.838	0.000	0.732	0.000				
	Aphis spp.	0.368	0.121	0.071	0.286	66.020	0.000	94.8%	
	B. tabaci	0.389	0.100	-0.264	0.702				
Orius sp.	Empoasca spp.	0.399	0.091	0.103	0.859				
-	P. solenopsis	-0.439	0.060	-0.080	0.001				
	N. viridula	0.942	0.000	0.707	0.000				
	Aphis spp.	0.729	0.000	0.287	0.015	9.230	0.001	69.6%	
	B. tabaci	-0.036	0.882	0.088	0.936				
Syrphus sp.	Empoasca spp.	-0.033	0.892	-0.113	0.903				
	P. solenopsis	0.315	0.189	0.064	0.048				
	N. viridula	0.621	0.005	0.264	0.006				
	Aphis spp.	0.600	0.007	0.291	0.042	5.060	0.009	53.0%	
	B. tabaci	-0.182	0.455	1.476	0.299				
A. aphidimyza	Empoasca spp.	-0.191	0.433	-1.255	0.249				
, ,	P. solenopsis	0.527	0.020	0.091	0.028				
	N. viridula	0.320	0.181	0.144	0.183				
	Aphis spp.	0.574	0.010	0.284	0.112	7.400	0.002	64.0%	
	B. tabaci	0.478	0.038	-1.359	0.453				
C. carnea	<i>Empoasca</i> spp.	0.489	0.034	1.288	0.399				
	P. solenopsis	-0.146	0.552	0.041	0.401				
	N. viridula	0.791	0.000	0.375	0.014				



Figure 1: Seasonal mean number of the predaceous insects on the four vegetable crops during 2022 at Kafr-Saad region Damietta Governorate.



Figure 2: Seasonal abundance of predatory insects in response to host plants (Cucumber, cowpea, tomato and sweet potato) at Kafr-Saad region Damietta Governorate during 2022.





Ata

Figure 3: Seasonal abundance of the coccinillid predator in response to prey densities on cucumber, cowpea, tomato and sweet potato plants during 2022.



Figure 4: Seasonal abundance of the *C. carnea* predator in response to prey densities on cucumber, cowpea, tomato and sweet potato plants during 2022.



Figure 5: Seasonal abundance of the *A. aphidimyza* & *Surphus* sp. predator in response to prey densities on cucumber, cowpea, tomato and sweet potato plants during 2022.



Figure 6: Seasonal abundance of the *Macroliphus* sp. & *Orius* sp.predator in response to prey densities on cucumber, cowpea, tomato and sweet potato plants during 2022 seasons.

دور الحشرات المفترسة في تنظيم تعداد الآفات الحشرية الثاقبة الماصة الرئيسية التي تهاجم بعض محاصيل الخضر

طارق السيد عطا قسم *وقاية النبات, كلية الزراعة, جامعة دمياط, دمياط, مصر.* * البريد الإلكتروني للباحث الرئيسي: drtarekata@du.edu.eg

الملخص العربي:

أجريت التجارب لدراسة دور الحشرات المفترسة في تنظيم مجاميع الآفات الحشرية الثاقبة الماصة على محاصيل الحضر في منطقة كفر سعد بمحافظة دمياط، مصر خلال موسم الزراعة الصيفى. أظهرت النتائج وجود تسعة مفترسات تنتمي إلى أربعة رتب حشرية هي ؛ غمدية الأجنحة، متباينة الأجنحة، ثنائية الأجنحة وشبكية الأجنحة على نباتات الخيار واللوبيا والطراطم والبطاطا الحلوة. وكانت مفترسات أبو العيد (أبو العيد أحد عشر نقطة *Coccinella* ثنائية الأجنحة وشبكية الأجنحة على نباتات الخيار واللوبيا والطراطم والبطاطا الحلوة. وكانت مفترسات أبو العيد (أبو العيد أحد عشر نقطة *Coccinella ثنائية الأجنحة وشبكية الأجنحة على نباتات الخيار واللوبيا والطراطم والبطاطا الحلوة. وكانت مفترسات أبو العيد (أبو العيد أحد عشر نقطة <i>Coccinella ثنائية الأجنحة وشبكية الأجنحة على نباتات الخيار واللوبيا والطراطم والبطاطا الحلوة. وكانت مفترسات أبو العيد (أبو العيد أحد عشر نقطة undecimpunctata دسلوكوروس (<i>Chilocorus nigritus ورفرة ع*لى العوائل النباتية المختبرة مقارنةً بكل من: بقة اوريس *Orius spp ورفوس (Chilocorus nigritus مواطول والوفوس Syrphus sp. ديابة المن وورة ع*لى العوائل النباتية المختبرة مقارنةً بكل من: بقة اوريس *Orius sp. محاصل الحور ورفو منات عائلة أبو العيد هي و 20.7 و 20.5 (من إج*الي عدد الحشرات المفترسة على محاصيل الخيار واللوبيا والطراط والبطاطا الحلوة على مفترسات عائلة أبو العيد ديابة السيرفس العد الأصغر بنسبة 5.4 و 20.5 *(من إج*الي عدد الحشرات المفترسة على مواليوا الحلوة على التوالي، بينا سجلت ذبابة السيرفس العد الأصغر بنسبة 5.4 و 20.6 *و 10.5 (من إج*الي أعداد الحشرات المفترسة على الحابرات المفترسة على الحاصرات المفترسة على العاصيل السابقة. فيا مفترسات عائلة أبو العيد ذبابة السيرفس العد الأصغر بنسبة 5.4 و 20.6 *و 20.6 و 20.6 ورفو ع*لى يتعلق بتفصيل الحشرات المفترسة للعوائل النباتية ، إتصح أن جميع المفترسات تفضل نباتات الخيار (450 مفترس/ 25 ورقة) تليا اللوبيا والوبيا والوبيا وروم 20.6 ورفقا و ووقة) والطراطم (114 مفترسة ملعوائل النباتية ، إتصح أن جميع المفترسات تفضل نباتات الخيار (450 مفترس/ 25 ورقة) تلقا بمرات المفترسة مملية بروم 20.6 ومفترس/ 25 ورقة) ووقة). وروتة معن الحاطم (111 مفترسة للعوائل النباتية

الكليات الاسترشادية: المفترسات الحشرية، محاصيل الخضر، الحشرات الثاقبة الماصة.